DECLARATION

I, ___Julio César Aguilar Rubido___

Translator, of the **CENTRO DE INGENIERIA GENETICA Y BIOTECNOLOGIA, Ave. 31 entre 158 y 190, Cubanacán, Playa, C. Habana 10600, Cuba,** do solemnly and sincerely declare that I am conversant with the English and Spanish languages and am a competent translator thereof, and that the annexed document is, to the best of my knowledge and belief, a complete, true and correct translation of the Cuban Patent Application 183/98 made in Cuba filed on December 12, 1998.

Date: April 2, 2001.

Signature: _____

REPÚBLICA DE CUBA

Lic. América N. Santos Riveras, Directora General de la **OFICINA CUBANA DE LA PROPIEDAD INDUSTRIAL**.

CERTIFICO: Que bajo el número ciento ochenta y tres del año mil novecientos noventa y ocho del Registro de Entrada, fue presentada en esta **OFICINA CUBANA DE LA PROPIEDAD INDUSTRIAL**, solicitud de Certificado de Autor de Invención por **FORMULACIONES CONTENIENDO PARTÍCULAS SEMEJANTES A VIRUS COMO INMUNOPOTENCIADORES POR VÍA MUCOSAL**, con fecha dos de diciembre de mil novecientos noventa y ocho, a las trece horas y treinta minutos pasado meridiano, por Mariela Vázquez Castillo, Representante, ciudadana cubana, a nombre y en representación del **CENTRO DE INGENIERÍA GENÉTICA Y BIOTECNOLOGÍA (CIGB)**, cuya invención fue creada por Julio César Aguilar Rubido; Daniel Octavio Palenzuela Gardon; Verena Lucila Muzio González; Gerardo Enrique Guillén Nieto; Eduardo Pentón Arias; Dagmara Pichardo Díaz; Enrique Iglesias Pérez.

ASIMISMO CERTIFICO: Que la mencionada solicitud de Certificado de Autor de Invención, se encuentra actualmente en tramitación.

TAMBIÉN CERTIFICO: Que la Memoria Descriptiva, Reivindicaciones y Dibujos que se acompañan, son exactamente iguales a las que obran en el expediente.

Y a petición de Mariela Vázquez Castillo, Representante, se expide la presente en la Ciudad de La Habana, República de Cuba, a los veintitrés días del mes de noviembre de mil novecientos noventa y nueve.

Lic. América N. Santos Riveras
Directora General

CUBAN OFFICE OF THE INDUSTRIAL PROPERTY

Dr. América N. Santos Riveras, Director of the **CUBAN OFFICE OF THE INDUSTRIAL PROPERTY.**

I **CERTIFY** that under the Entry Registry Number hundred eighty three of the year nineteen ninety eight, it was presented before the **CUBAN OFFICE OF THE INDUSTRIAL PROPERTY** the Certificate of Authorship: **FORMULATIONS CONTAINING VIRUS-LIKE PARTICLES AS MUCOSAL IMMUNOENHANCERS,** dated on December two, nineteen ninety eight, at thirteen and thirty pm, by Mariela Vázquez Castillo, Representative Agent, Cuban citizen, on behalf and representing the Centro de Ingeniería Genética y Biotecnología (CIGB), whose invention was created by Julio César Aguilar Rubido, Daniel Octavio Palenzuela Gardon; Verena Lucila Muzio González, Gerardo Enrique Guillén Nieto, Eduardo Pentón Arias, Dagmara Pichardo Díaz, and Enrique Iglesias Pérez.

LIKEWISE, I CERTIFY: that the above-mentioned Certificate of Author of the Invention is currently under process.

I ALSO CERTIFY: that the Disclosure of the Invention, the Claims and the Figures, which are enclosed, are equal to those that are included in the dossier.

At the request of Mariela Vázquez Castillo, Representative Agent, this certification is released in Havana City, Republic of Cuba, on twenties three days of the month of November, nineteen ninety nine.

Dr. América N. Santos Riveras
General Director

DESCRIPTIVE MEMORY

FORMULATIONS CONTAINING VIRUS-LIKE PARTICLES AS MUCOSAL IMMUNOENHANCERS.

The present invention is related to the branch of medicine, particularly with the use of new vaccine immunoenhancing strategies. In this case, the adjuvant is a virus-like particle (VLP), which at the same time constitutes an antigen of interest in the formulation. The adjuvant mechanism is based on the positive effect of one antigen on others or on the synergic interaction between the antigens of the formulation.

The technical objective pursued with the present invention is, precisely, the development of formulations capable of enhancing the immune response to antigens administered through mucosal routes, minimizing the number of components in the formulation. The enhancing activity is supported by the interaction between particles at the mucosal level, generating systemic as well as mucosal immunity. Furthermore, the development of combined vaccines to the mucosal route taking as a central antigen the HBsAg, increased the immune response to one or more of coadministered antigens. The obvious advantage is the elimination of all other element or compounds different from the antigen of interest and the use of a different route. We consider that this is the basis or nucleus to develop combined vaccines for a mucosal use.

HBcAg is an extremely immunogenic antigen during the Hepatitis B Virus (HBV) infection or after immunization. In many HBV chronic patients, this is the only antigen capable of inducing an immune response. It can even induce an immune response in mice in nanogram quantities. Recently, a few structural studies have demonstrated some important characteristics explaining its potent immunogenicity. HBcAg specifically binds membrane immunoglobulin receptors in a large number of resting B cells in mice, which is sufficient to induce costimulatory B7-1 and B7-2 molecules. In this way, non-sensitized B cells, specific for HBcAg can uptake, process and present HBcAg peptides to naive T cells *in vivo* and to T cells hybridoma *in vitro,* approximately 10^5 times more efficiently than macrophages and dendritic cells. This

structure-function relationship explains the great immunogenicity of HBcAg (Milich, D.R. *et al.* 1997 Proc. Natl. Acad. Sci USA Dec23; 94(26): 14648-53). Serologic and biochemical studies indicate that the resolution of HBV acute infection occurs in the context of an efficient cell-mediated immune response, while the chronic infection is characterized by a poor and undetectable cell-mediated immune response and a "relatively efficient" humoral response.

The humoral immunity and the cell-mediated immunity are regulated by different groups of helper T cells. Factors influencing the induction in mice of a Th1 or Th2 response to the HBV antigens (HBcAg/HBeAg) revealed that this balance was influenced (1) by the antigen structure (HBcAg is a particulated structure and HBeAg is not; (2) the major histocompatibility complex (MHC) of the host and the T cell antigens which are recognized; (3) the cross regulation between Th1 and Th2 cells; (4) the T cell tolerance, which is more complete for Th1 than for Th2 cells; (5) the activity of secreted HBeAg that preferentially delete Th1 cells (6) the treatment with cytokines, used to modulate *in vivo* the response toward Th1 or Th2 cells. This balance Th1/Th2 is relevant to the acute or chronic course of the HBV infection. Th2 cells preferentially evade the induction of tolerance compared with Th1. As HBeAg acts as a tolerogen during HBV vertical transmission, deleting Th1 cells, the predominance of Th2 specific cells for HBeAg could influence in the initiation and maintainance of a chronic carrier state. In this case the cytokine therapy endowed to modulate the response towards Th1, could be benefitious in the treatment of HBV chronic infection (Milich, D.R. 1997 J. Viral. Hepat.; 4 Suppl 2: 48-59).

The effect of HBeAg circulation on HBcAg Th1 specific T cells was examined by transferring HBeAg/HBcAg specific T cells to double (HBeAg and HBcAg) transgenic mice. The presence of serum HBeAg eliminated the Th1 mediated response against HBcAg and changed the balance to the Th2 phenotype. This result suggest that, in the context of the hepatitis B infection, circulating HBeAg has the potential to preferentially eliminate inflammatory specific Th1 cells needed for viral clearance, promoting the persistency of HBV (Milich-DR *et al.* 1998 J-Immunol. Feb 15; 160(4): 2013-21).

It is known that antibodies against HBcAg are present since the beginning of the infection and reach high concentrations in sera of HBV chronically infected

patients, but these antibodies are not protective. Antibodies passively transmitted to newborn children by chronic carrier mothers, do not protect children of infection. (Beasley et al. 1977. American Journal of Epidemiology 105: 914-918). However, it has been demonstrated that immunizing chimps with HBcAg partially or completely protected them from HBV infection (Iwarson, S. et al. 1985 Gastroenterology 88: 763-767; Murray, K. et al. 1987 Journal of Medical Virology 23: 101-107). In Iwarson's study, three chimps were completely protected. After challenge with HBV, antibody levels against HBcAg and HBeAg increased but only one chimp seroconverted against HBsAg. In Murray's study, 2 out of 4 immunized chimps showed a low level of viral replication after challenge, HBsAg was detectable in sera for 2 or 3 weeks, and after that they developed an anti-HBsAg antibody response. It was hypothesized that the incomplete protection could be due to the low immune response in vaccinated animals without adjuvant.

After immunizing with woodchucks hepatitis core antigen (WHcAg) in Freund Complete Adjuvant (ACF), it was possible to protect woodchucks from challenge with the virus (WHV) without signs of infection detectable antibodies against the surface protein (WHsAg). Although the hypothesis that T helper anti-nucleocapsid immune response could enhance undetectable antibodies against the surface antigen can not be discarded, the cytotoxic activity was considered as the main responsible of protection (Roos, S. et al. 1989 J. Gen. Virol. 70, 2087-2095). In a second study using woodchucks the role of HBcAg and WHcAg in protection was determined as well as the possible mechanism. Animals were immunized with WHcAg and HBcAg and afterwards challenged using a high dose of WHV. In this experiment it was found that WHcAg is a protecting antigen there is a cross protection because 4 out of 6 woodchucks immunized with HBcAg were protected from the challenge. Both antigens generated a high antibody titer with a cross reactivity lower than 1%, confirming previous reports of protection using internal hepatitis B virus antigens. Since dominant B epitopes of both antigens do not appear to be conserved, this result also demonstrated that antibodies directed against core antigens are not important for protection. Woodchucks immunized with WHcAg/HBcAg reacted with a rapid response of serum antibodies against

surface proteins after challenge with WHV, indicating an increased helper T cell response as a potential mechanism of protection after immunization with an internal antigen of HBV/WHV. (Schodel-F *et al.* Vaccine. 1993; 11(6): 624-8) Transfection of established cell lines from BALB/C mice hepatocytes with dimeric HBV DNA (ML lines) resulted in the expression of HBV antigens in these lines. The adoptive transference of spleen cells of BALB/c mice immunized with ML-1.1 cells expressing HBsAg as well as HBcAg, caused a regression of tumours cells expressing the corresponding antigens in athymic mice. Furthermore, the transfer of spleen cells of BALB/c mice immunized with HBsAg or HBcAg also caused tumoral regression. These results demonstrated that surface and nucleocapsid antigens could induce immunity capable of rejecting the hepatocellular carcinoma *in vivo* (Chen, S.H. *et al.* 1993 Cancer-Res. Oct 1; 53(19): 4648-51).

Therapeutic vaccines based on specific nucleocapsid epitopes for human HLA are being assayed in phase II/III studies (Liaw, Y.F. 1997 J.Gastroenterol. Hepatol. Oct, 12 (9-10): S227-35).

HBcAg has been demonstrated to be a very good carrier. HBcAg represents a highly immunogenic antigen in human and animal models. HBcAg activates directly B cells and generates strong T cell responses, furthermore, the efficient processing and presentation of HBcAg by the antigen presenting cells makes it the ideal carrier molecule. Hence a large number of epitopes has been chemically linked or genetically fused to the HBcAg molecule to successfully increase their immunogenicity. Expression vectors has been designed in bacteria to enable the insertion of heterologous B cell epitopes in different positions inside the particles of HBcAg and the efficient purification of hybrid particles.

Positional studies of B cell epitopes demonstrated that internal insertions by the amino acid 80 continue to be immunodominant, permitting an increase in the production of antibodies as compared to other fusion proteins. Immunogenicity studies have been performed with experimental challenge in different systems. For example, a peptide from *Plasmodium berghei* Circunsporozoite was inserted in this site and the purified hybrid particle HBcAg/CS was highly immunogenic and protected 100% of challenged mice

against malaria. Aimed to the development of oral vaccines, attenuated avirulent Salmonella strains have been used to introduce genes coding for hybrid particles of HBcAg (Milich, D.R. *et al.* 1995 Ann. N.Y. Acad. Sci. May 31; 754: 187-201).

In conclusion, apart from the relationship between HBcAg and protection, total or partially evidenced in chimps or indirectly referred by the experiments with WHcAg, this protein has a number of properties that makes it unique. HBcAg behaves as a T dependent as well as a T independent antigen (Milich, D.R. *et al.* 1986 Science 234, 1398-1401), it is very immunogenic, even without the help of adjuvants and its inoculation preferentially sensitizes Th1 cells (Milich, D.R. et al. 1997, J. Virol. 71, 2192-2201). HBcAg is a very efficient carrier protein (Schödel, F. *et al.* 1992 J. Virol. 66: 106-114; Milich-DR *et al.* 1995 Ann-N-Y-Acad-Sci. May 31; 754: 187-20) and Th HBcAg specific cells mediate the antibody response against HBcAg as well as anti-HBsAg (Milich, D.R. et al. 1987 Nature (London) 329: 547-549). These immunologic characteristics are unique for the particulated HBcAg and do not apply to the non-particulated form of the antigen, the HBeAg (Milich, D.R. *et al.* 1997 Proc. Natl. Acad. Sci USA Dec 23; 94(26): 14648-53).

Detailed description of the invention

In the present invention it is reported for the first time a vaccine formulation having as main compounds: HBsAg and HBcAg. Derived compounds from both antigens can also be contained. These derived compounds can be obtained by genetic engineering, chemical conjugation or adsorption, in adequate proportions. Other compounds may be introduced as stabilizers and preservatives.

The novelty of HBsAg/HBcAg formulation is linked to the anti HBsAg enhancing effect generated when HBsAg is mixed with HBcAg. Both antigens are compounds of HBV and hence, the role of the adjuvant is taken by other viral antigen attractive *per se* as a vaccine antigen, becoming a vaccine formulation with a wider anti-hepatitis B immune response spectrum. Other formulations of nucleocapsid antigens combined with surface antigens, for example the formulation HBsAg/Virus like particle of Human Papilloma Virus and extended to other viral antigens, results in an increase in titers against

both antigens. After mixing HBsAg with other antigens, an increase in the immunogenicity over other coinoculated antigens could be shown, evidencing a synergic effect produced by the combination X + HBsAg through the nasal route. In general, these results enables the generation of HBsAg mucosal combined vaccines, and enables the use of the positive interactions between VLP, considering VLP as organized proteic or lipoproteic structures, resembling viruses, with a size of nanometers.

In the case of the formulation containing HBsAg and HBcAg, we can obtain a superior product as compared to the single HBsAg commercially available vaccine because:

-It is possible to obtain a wider spectrum of immune response generated by HBcAg regarded as an important antigen *per se* in anti-VHB protection. Furthermore, the IgG seric levels anti-HBsAg reached by mucosal inoculation is as intense as the one obtained with the systemic inoculation in alum.

-The route of inoculation offers special advantages such as: systemic and mucosal immunity at the same time, the elimination of strong quality controls such as sterility and pyrogens as well as the high prices of injected vaccines, the related toxicity.

-It is possible and obvious to include antigens derived from HBsAg and HBcAg, using their carrier capacity in the context of this formulation introducing in this way important homologous or heterologous antigens.

-The toxic effect generated by alum-based vaccines and the toxic effects of adjuvant injection can be avoided because the antigen number 2 is at the same time the adjuvant.

-It is possible to use the initial HBsAg + HBcAg formulation as a nucleus of combined vaccines.

-It is possible to immunize non responders to the surface antigen and immunodepressed patients using this preparation, due to the inoculation route and the introduction of the nucleocapsid antigen regarded as a protective antigen *per se*.

-The characteristics of this formulation make it an ideal formulation for therapeutic use.

In the second place, nucleocapsid antigens, favours the increase of the immunogenicity of coinoculated antigens. We found a great simplicity of resulting formulations and, at the same time, the increased valence of these potential vaccines with a minimal number of antigens due to the possibility of avoiding the use of adjuvants, which are *per se* non-interesting antigens for protection. In this way very reduced combinations can be obtained if desired, for single or combined vaccines.

In the third place, it is possible to generate combined vaccines having as a nucleus the HBsAg whose immunoenhancing effect on other coinoculated antigens is demonstrated in the example 4. The advantages of these formulations are based on the effective association of HBsAg, as a central antigen of the anti HBV vaccine, with other antigens, with a demonstrated synergic effect in the generated response for both antigens. This fact, not only has the attraction of previously described variants but it also makes the HBsAg, -protecting antigen for a widely distributed world disease- the central antigen of combined formulations.

In general terms, compared to other mucosal vaccines, it is possible to detach the following advantages:

-The 'adjuvation' process −mixing antigens- does not require the adsorption of antigens, and the quantity of the HBcAg antigen is at similar levels of HBsAg.

-The filtration as a sterilizing process can be used due to the small size of the particles, while other strategies and adjuvants over $0.2\mu m$ can not be sterilized in this way.

-The simplicity of the production process for HBcAg makes it a very cheap antigen as compared to other adjuvants.

The formulations object of the present invention may present volumes from 0.01 until 10mL, depending of the inoculation route and the species to immunize. The antigen doses vary in a range of 0.001 to 1mg.

EXAMPLES OF PERFORMANCE

Example 1

With the aim of evaluating the immunogenicity of HBcAg through the nasal route, 3 groups of 8 female BALB/c mice were inoculated with a dose of 10μg of HBcAg in all cases. The first group was inoculated with HBcAg in acemannan (CIGB, La Habana) 3 mg/mL (dry weight), adjuvant previously used to increase the immunogenicity of particulated systems through the nasal route. The second group was inoculated with HBcAg in phosphate-saline (PBS) buffer. Group 3 was injected subcutaneously with the antigen in alum and used as a control group for systemic inoculation. The schedule was followed of inoculations on days 0, 14, 28 and the extraction was done on day 42. The antibody response was quantified by immunoenzymatic assay (ELISA) to determine the IgG antibodies against HBcAg in sera.

The Student's t test was performed to analyze statistically the results, $p < 0.05$ was considered a significant difference.

It was demonstrated that, with the use of acemannan it was impossible to increase the anti-HBcAg antibody immune response. The antigen in PBS generated an immune response of a similar intensity to that obtained using acemannan (Fig. 1). The responses after nasal inoculation, in acemannan or in PBS, were similar to the response obtained using alum systemically. In conclusion, HBcAg can be used through the nasal route with a high immunogenicity.

Example 2

With the aim of demonstrating the immunoenhancing activity of HBcAg on HBsAg when both are mixed and inoculated through the nasal route, 4 groups of 8 female BALB/c mice were assayed. A two inoculations schedule was carried out. The inoculations were on days 0 and 14. The extraction was on day 21. The group 1 was inoculated with 10μg of HBsAg in PBS, group 2 with 10μg of HBsAg in acemannan (CIGB, La Habana) 3mg/mL (dry weight), group 3 with 10μg of HBsAg and 10μg of HBcAg. Group 4 was used as a systemic control, inoculating subcutaneously 10μg of HBsAg in alum (Fig. 2). The Student t test was performed to analyse statistically the results, $p < 0.05$ was considered a significant difference.

From this experiment we concluded that it is possible to enhance the anti HBsAg immune response with the inoculation through the nasal route of HBsAg and HBcAg. The anti HBsAg response was significantly superior as compared to the group when the HBsAg was inoculated in PBS and similar to that reached by the group inoculated in acemannan. The systemic inoculation of HBsAg in alum did not differ significantly from the groups inoculated with acemannan through the nasal route.

Example 3

With the aim of studying the enhancing effect of HBcAg at different doses in the murine model, 6 groups of 6 female BALB/c mice were selected. The schedule had three inoculations (days 0, 14 and 28) and two extractions (days 26 and 42). The assayed groups corresponded with: (1) HBsAg 5 μg in PBS; (2, 3 y 4) HBsAg 5 μg with 5, 10 y 20μg of HBcAg respectively, (5) HBsAg 5μg in acemannan 3mg/mL (dry weight) and (6) HBsAg 5μg in alum 0.5mg/mL. All groups except 6 were inoculated nasally. Group 6 was inoculated intramuscularly.

The Student's t test was used to analyze statistically the results, $p < 0.05$ was considered a significant difference.

In this experiment we concluded again that it is possible to enhance the anti-HBsAg response with the coinoculation of HBsAg and HBcAg. The serum IgG response for the three immunized groups with both antigens was significantly higher to that obtained by inoculation of HBsAg in PBS and similar to that attained by the group inoculated in acemannan. We have demonstrated previously that acemannan increased the titers to levels similar to that obtained by cholera toxin in mice. Titers obtained by systemic inoculation in alum did not differ from that of acemannan and HBcAg/HBsAg groups by the nasal route. Although group 4 anti-HBsAg antibody response decreased as compared to group 3, the difference was due to a double increase in the HBcAg dose in group 4. This increase might reduce the possibilities of HBsAg to penetrate mucosa.

Example 4

Different antigens were employed with the aim of studying the interaction of virus-like particles of Human Papilloma Virus 16 (VLP del VPH 16), HBsAg

and HBcAg. Were immunized 8 groups of 6 female BALB/c mice with a schedule based in inoculations on days 0, 14 and the extraction 7 days after second inoculation.

Comparing antibody titers against HBsAg, the response of acemannan formulation (group 6) has the same intensity to the HBcAg/HBsAg formulation (group 7) respectively. This is the third time that we demonstrate the enhancing effect of HBcAg.

From this experiment we also concluded that neither acemannan nor HBcAg enhanced antibody responses against VLP of Human Papilloma Virus (HPV), represented as groups 4, 5 and 8 in the third graphic. Statistical analysis using Student's t test ($p < 0.05$ was considered a significant difference) did not show any difference between these groups.

Analysing the response against HBcAg in the group 5, where HBcAg and VLP of HPV were inoculated, low levels of antibody titers against HBcAg could be demonstrated as compared to group 7, where HBcAg was introduced along with HBsAg. Perhaps, the presence of these two particles antagonizes at the mucosal level. However, in group 2, high anti HBcAg and anti VLP of HPV could be achieved with the addition of HBsAg, being significantly higher the increase in these responses as compared to group 5 and do not differ from anti HBcAg response of group 7 (along with HBsAg). Hence we could realize a positive interaction between HBsAg and core antigens and a negative interaction between VLPs and HBcAg. The enhancing effect at mucosal level can occur in both senses, enabling the design of combined vaccines having as a nucleus HBsAg or the HBsAg/HBcAg combination.

HBsAg effect on group 2 not only enhanced the response against HBcAg, but it also enhanced the antibody response against the VLP of HPV. The same effect can be appreciated comparing the response against VLP between groups 1, 2 and 3 with group 8 where VLP were inoculated in PBS. Groups 1, 2 and 3 had statistically similar antibody levels, all of them higher than the group 8 level.

Comparing group 1 and 3: where the polysaccharide is added to the mixture (group 1) or not (group 3), there are no significant responses regarding to

anti HBsAg titers. This result evidenced the enhancing effect of VLP of HPV on HBsAg immunogenicity.

These results support the use of combined formulations through nasal routes with HBsAg as a central immunoenhancing antigen. For example, the simple mixture of HBsAg and HPV VLP is very attractive and makes real the possibility of introducing more antigens, enhanced by the interaction with HBsAg.

These antigens, inoculated through the nasal route, as an advantage have the posibility of forming more complex formulations, not affected in antibody titers with the introduction of new antigens in the formulation (example: the introduction of VLP of HPV to HBcAg and HBsAg in the group 2), compared with combined formulations of two antigens as HBcAg/HBsAg or single antigen formulations as HBsAg/acemannan.

DESCRIPTION OF FIGURES

Figure 1. Three doses schedule (days 0, 14 and 28). Extraction was performed on day 42. Groups 1 and 2 were inoculated with 50µL through the nasal route. Group 3 was inoculated subcutaneously with 100µL.

Figure 2. Two doses schedule (days 0 and 14). Extraction was performed on day 21. Groups 1, 2 and 3 were inoculated with 50µL through the nasal route. Group 4 was inoculated subcutaneously with 100µL.

Figure 3. Three doses schedule (days 0, 14 and 28). Extraction was performed on day 26. Groups 1, 2, 3, 4 y 5 were inoculated through the nasal route. Group 6 was inoculated intramuscularly with 100µL.

Figure 4. Two doses schedule (days 0 and 14). Extraction was performed on day 26. All groups were inoculated nasally with 50µL. The composition of experimental groups is shown in table added to the figure.

Lic. Mariela Vázquez Castillo
Representante Legal, CIGB

CLAIMS

FORMULATIONS CONTAINING VIRUS-LIKE PARTICLES AS MUCOSAL IMMUNOENHANCERS.

1. A pharmaceutical formulation wherein its main components are a) the Hepatitis B surface antigen and b) the Hepatitis B nucleocapside antigen, other components are preservatives and stabilizers.

2. A pharmaceutical formulation wherein its main components are a) the Hepatitis B surface antigen and/or a derivative of HBsAg and b) the Hepatitis B nucleocapside antigen and/or a derivative of HBcAg, other components are preservatives and stabilizers.

3. A formulation according to claim 2, wherein the HBsAg derivative and/or HBcAg derivative constitutes an antigen or portion of an antigen, of any nature, genetically inserted, chemically coupled or adsorbed to the structure of one or both antigens.

4. A formulation according to claims 1, 2 and 3, wherein the addition of homologous or heterologous antigen is enhanced by the addition of HBcAg.

5. A formulation according to claims 2, 3 y 4, wherein the antigen can be included inside of the structure of any of the particulated antigens.

6. A formulation according to claim 5, wherein the antigen trapped is a nucleic acid.

7. A pharmaceutical formulation according to claims 1, 2, 3, 4, 5 and 6 for mucosal use.

8. A pharmaceutical formulation according to claims 1, 2, 3, 4, 5 and 6 for systemic use.

9. A pharmaceutical formulation according to claims 1-7 as a therapeutic vaccine.

10. A pharmaceutical formulation according to claims 1-7 as a preventive vaccine.

11. A pharmaceutical formulation according to claims 1, 2, 3, 4 and 5 containing at least two antigens, one of them the HBsAg and the other, a viral nucleocapsid, for mucosal use.

12. A formulation for mucosal administration according to claim 11, containing as enhancing element a viral nucleocapsid.

13. A formulation for systemic administration according to claim 11, containing as enhancing element a viral nucleocapsid.

14. A formulation according to claim 11 containing as immunoenhancer the antigen of nucleocapside of the HBV in a simple mixture without covalent binding.

15. A formulation for mucosal immunization of any antigen mixed with HBsAg, using the enhancing effect of HBsAg.

16. A combined vaccine for mucosal use using as a central antigen de Hepatitis B surface antigen.

Lic. Mariela Vázquez Castillo
Representante Legal, CIGB

First schedule

1- 10μg HBcAg / acemannan 3mg/mL   IN

2- 10μg HBcAg / PBS 1X   IN

3- 10μg HBcAg / alum 0.5mg/mL   SC

Fig. 1

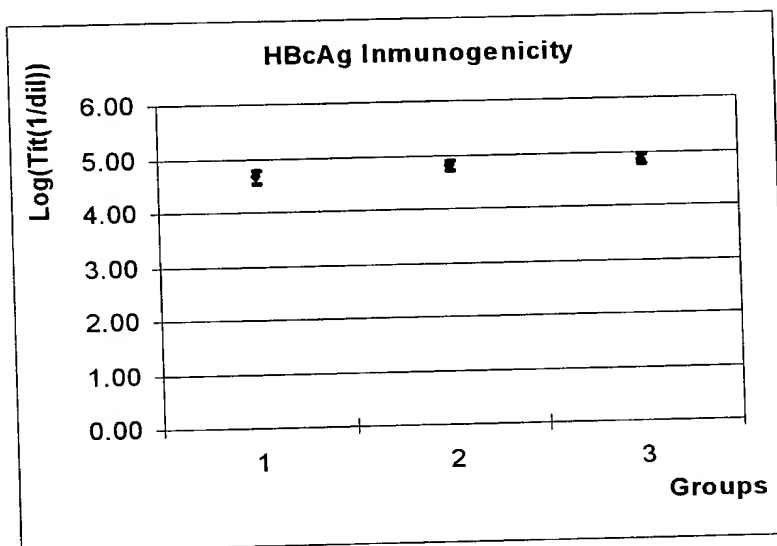**Second Schedule**

1-10μg HBsAg/ PBS 1X   IN

2-10μg HBsAg/ acemannan 3mg/mL   IN

3-10μg HBsAg/ 10μg HBcAg / PBS 1X   IN
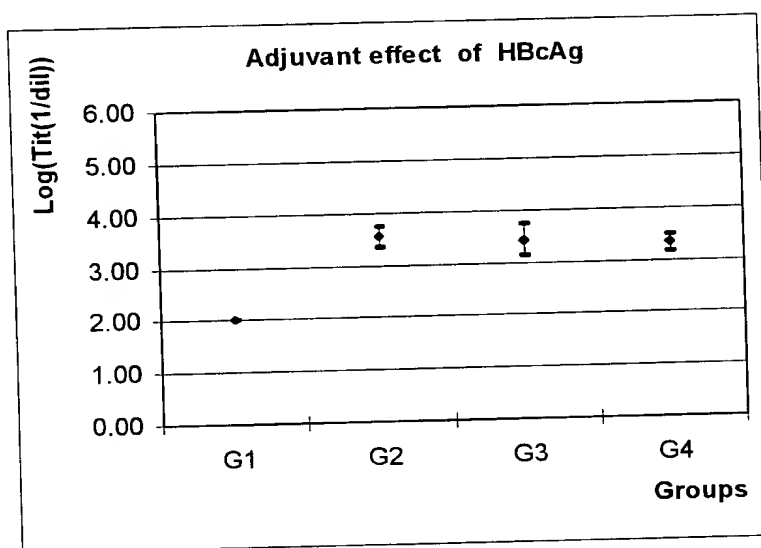
4-10μg   HBsAg/ Alum 0.5mg/mL   SC

Fig. 2

Third schedule

1- 5µg HBsAg / PBS 1X   IN

2- 5µg HBsAg / 5µg HBcAg   IN

3- 5µg HBsAg / 10µg HBcAg   IN

4- 5µg HBsAg / 20µg HBcAg   IN

5- 5µg HBsAg / acemannan 3mg/mL   IN

6- 5µg HBsAg / alum 0.5mg/mL   IM

Adjuvant effect of HBcAg
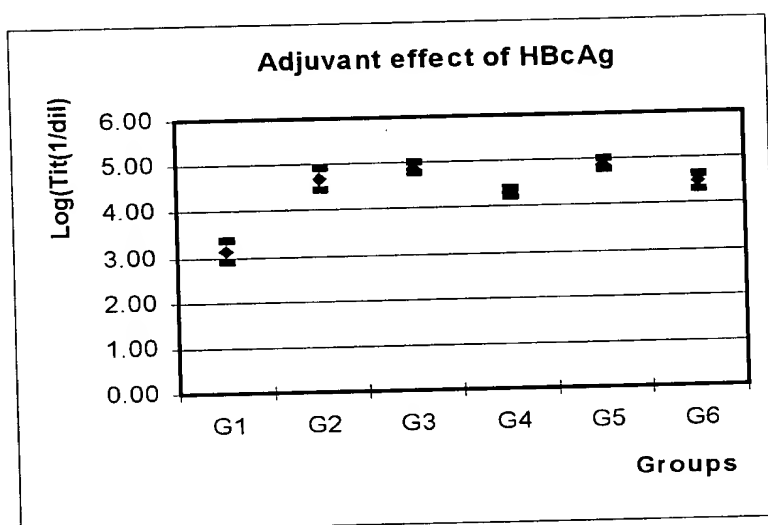
Fig. 3

Fourth Schedule: Synergism at mucosal level.

	G1	G2	G3	G4	G5	G6	G7	G8
Acemannan 3mg/mL	X			X	X		X	
HBcAg 5µg/dose		X			X		X	
HBsAg 5µg/dose	X	X	X			X	X	
VLP /HPV 5µg/dose	X	X	X	X	X			X

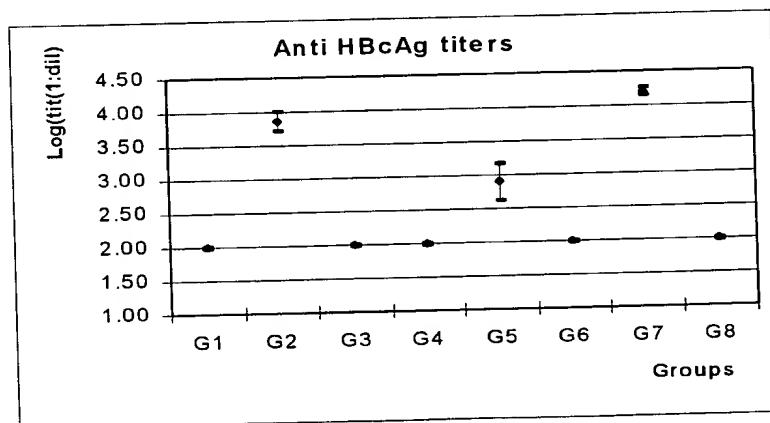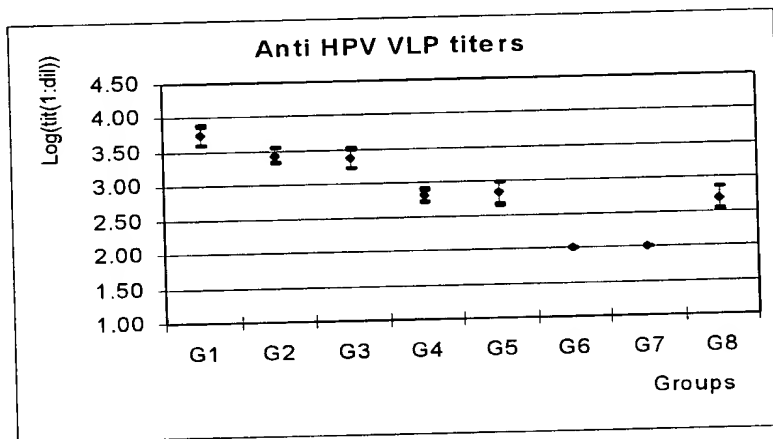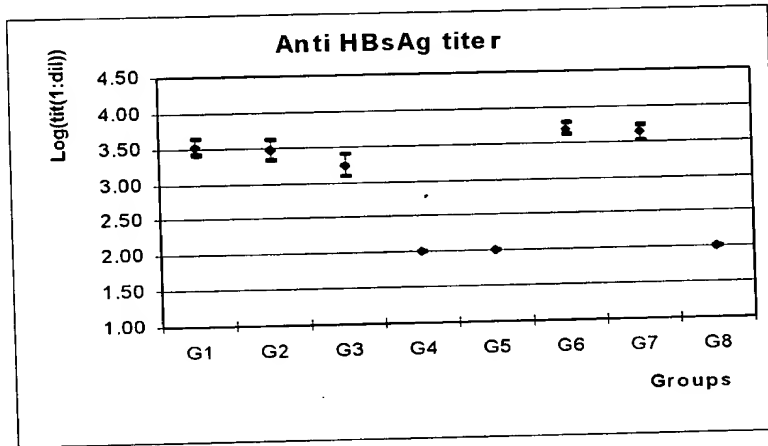**Anti HBsAg titer**

Anti HPV VLP titers

Anti HBcAg titers

Fig. 4 Composition, per groups, in the upper part of the figure.